

# Introduction to RBM package

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## 1 Overview

This document provides an introduction to the RBM package. The RBM package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the RBM package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

## 2 Getting started

The RBM package can be installed and loaded through the following R code.  
Install the RBM package with:

```
> source("http://bioconductor.org/biocLite.R")
> biocLite("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

## 3 RBM\_T and RBM\_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The  $p$ -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```
[1] 14
```

```

> which(myresult$permutation_p<=0.05)

[1] 3 43 57 69 100 105 166 321 460 490 759 891 925 957

> sum(myresult$bootstrap_p<=0.05)

[1] 3

> which(myresult$bootstrap_p<=0.05)

[1] 23 315 851

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 0

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 13

> which(myresult2$bootstrap_p<=0.05)

[1] 50 166 295 498 511 560 710 728 804 874 903 920 931

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the RBM\_F function: normdata\_F simulates a standardized gene expression data and unifdata\_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1  3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p   3000   -none-  numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 70

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 69

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 72

> which(myresult_F$permutation_p[, 1]<=0.05)

[1] 1 3 14 19 21 34 46 76 79 100 129 139 154 172 174 181 191 197 208
[20] 217 249 252 260 285 324 329 331 356 366 376 383 388 424 465 478 487 496 509
[39] 512 528 564 568 592 608 614 620 659 669 671 709 726 732 741 742 743 767 784
[58] 791 807 813 819 867 875 889 905 913 928 958 979 995

> which(myresult_F$permutation_p[, 2]<=0.05)

[1] 1 3 14 19 46 74 76 79 86 100 119 129 139 153 154 174 181 197 208
[20] 217 243 249 252 285 288 331 366 376 383 409 424 431 465 487 496 512 528 564
[39] 568 587 592 608 614 620 659 669 671 709 726 732 741 742 743 784 791 807 813
[58] 826 867 875 889 905 906 928 958 963 979 990 995

> which(myresult_F$permutation_p[, 3]<=0.05)

[1] 1 3 14 19 74 76 79 100 119 120 129 139 141 153 154 174 181 195 197
[20] 208 217 249 252 260 285 331 356 383 424 478 487 496 509 512 528 549 560 564
[39] 568 592 595 608 614 620 659 669 671 726 741 742 743 767 771 782 784 791 807
[58] 812 813 843 867 875 889 905 906 928 932 945 958 979 990 995

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

```

```

[1] 19

> con2_adj_p <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adj_p<=0.05/3)

[1] 12

> con3_adj_p <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adj_p<=0.05/3)

[1] 17

> which(con2_adj_p<=0.05/3)

[1] 3 19 197 285 331 564 608 659 671 791 807 995

> which(con3_adj_p<=0.05/3)

[1] 1 3 19 197 285 424 564 608 671 791 807 889 905 928 958 990 995

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

      Length Class  Mode
ordfit_t      3000  -none- numeric
ordfit_pvalue 3000  -none- numeric
ordfit_beta1  3000  -none- numeric
permutation_p 3000  -none- numeric
bootstrap_p   3000  -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 56

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 73

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 68

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

```

```

[1] 8 50 61 65 85 94 119 123 178 191 199 203 206 220 225 227 266 321 325
[20] 347 358 376 394 417 458 503 506 516 519 539 551 558 565 613 633 644 648 664
[39] 679 693 741 745 755 771 806 816 829 840 867 882 902 930 954 964 967 988

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 8 15 50 65 85 101 111 119 123 178 187 191 199 203 206 220 225 235 256
[20] 260 266 321 325 347 349 358 376 394 405 417 458 460 476 503 506 507 510 516
[39] 519 539 545 551 557 558 613 633 644 648 664 666 679 693 741 745 755 771 806
[58] 816 829 837 840 867 878 882 888 902 907 930 950 964 967 988 993

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 5 8 15 50 61 65 71 97 119 123 156 191 199 203 206 220 225 227 235
[20] 242 266 321 323 325 339 347 349 358 367 394 405 417 458 460 469 503 506 511
[39] 516 519 539 545 551 558 613 633 644 648 664 666 679 693 697 741 755 771 806
[58] 813 816 829 867 878 882 902 930 964 967 988

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 8

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 9

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 15

```

## 4 Ovarian cancer methylation example using the RBM\_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM\_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM\_T function and presenting the results for further validation and investigations.

```

> system.file("data", package = "RBM")

[1] "E:/biocbld/bbs-3.1-bioc/tmpdir/Rtmpa2cQH0/Rinst65c898746e/RBM/data"

> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)

      IlmnID      case1      case2      control1
cg00000292: 1  Min.   :0.01058  Min.   :0.01138  Min.   :0.009103
cg00002426: 1  1st Qu.:0.04111  1st Qu.:0.04290  1st Qu.:0.041543
cg00003994: 1  Median :0.08284  Median :0.10438  Median :0.087042
cg00005847: 1  Mean    :0.27397  Mean    :0.29086  Mean    :0.283729
cg00006414: 1  3rd Qu.:0.52135  3rd Qu.:0.54436  3rd Qu.:0.558575
cg00007981: 1  Max.    :0.97069  Max.    :0.96901  Max.    :0.970155
(Other)      :994
      control2      case3      case4      control3
Min.   :0.01019  Min.   :0.01108  Min.   :0.009753  Min.   :0.01278
1st Qu.:0.04092  1st Qu.:0.04059  1st Qu.:0.041818  1st Qu.:0.04260
Median :0.09042  Median :0.08527  Median :0.092807  Median :0.09362
Mean    :0.28508  Mean    :0.28482  Mean    :0.283113  Mean    :0.27563
3rd Qu.:0.57502  3rd Qu.:0.57300  3rd Qu.:0.558211  3rd Qu.:0.52240
Max.    :0.96658  Max.    :0.97516  Max.    :0.963620  Max.    :0.95974
      NA's      :1      NA's      :1
      control4
Min.   :0.01357
1st Qu.:0.04387
Median :0.09282
Mean    :0.28679
3rd Qu.:0.57217
Max.    :0.96268

> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)

      Length Class  Mode
ordfit_t      1000  -none- numeric
ordfit_pvalue 1000  -none- numeric
ordfit_beta0   1000  -none- numeric
ordfit_beta1   1000  -none- numeric
permutation_p 1000  -none- numeric
bootstrap_p    1000  -none- numeric

> sum(diff_results$ordfit_pvalue<=0.05)

[1] 31

```

```

> sum(diff_results$permutation_p<=0.05)

[1] 70

> sum(diff_results$bootstrap_p<=0.05)

[1] 41

> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adj_p<=0.05)

[1] 0

> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adj_p<=0.05)

[1] 13

> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adj_p<=0.05)

[1] 5

> diff_list_perm <- which(perm_adj_p<=0.05)
> diff_list_boot <- which(boot_adj_p<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t[diff_list_boot, ])
> print(sig_results_perm)

```

	IlmnID	case1	case2	control1	control2	case3
66	cg00059424	0.02742616	0.02554150	0.03049395	0.02910234	0.02547771
129	cg00121158	0.03045297	0.02728770	0.03573820	0.03316130	0.02853104
193	cg00177923	0.04979515	0.04607616	0.04060709	0.04200000	0.04504412
245	cg00224508	0.04479948	0.04477529	0.04152814	0.04189373	0.04208405
432	cg00419564	0.03638860	0.03661916	0.04101457	0.04065540	0.03283922
454	cg00436282	0.79300120	0.79687150	0.78143640	0.77336780	0.81890080
460	cg00445824	0.14782870	0.16655800	0.14393210	0.13479670	0.20038750
601	cg00577464	0.02668995	0.04306255	0.02710953	0.01929375	0.02770019
660	cg00634577	0.03182804	0.03432180	0.03525499	0.03612398	0.03384897
690	cg00661202	0.01639344	0.01586308	0.01876500	0.02097005	0.01490915
764	cg00730260	0.90471270	0.90207400	0.91002680	0.91258610	0.90575890
772	cg00743372	0.03922780	0.03499011	0.02187972	0.02568053	0.02796053
815	cg00792849	0.12674200	0.17255180	0.12769620	0.11480600	0.15856800
	case4	control3	control4	diff_results\$ordfit_t[diff_list_perm]		
66	0.02523713	0.04478458	0.03391813			-2.203752
129	0.02993313	0.03318921	0.03345607			-2.118418
193	0.04416582	0.03535173	0.04315951			2.080942
245	0.05731476	0.03775905	0.03955271			1.811314



432	0.03934095	0.04413387	0.04462037	-2.407553
454	0.81115150	0.67173700	0.77315940	2.213478
460	0.16185300	0.11630830	0.13912630	2.993440
601	0.03204772	0.02634007	0.02547999	1.786901
660	0.03062112	0.03502489	0.03710039	-1.402274
690	0.01764273	0.01847447	0.01803320	-1.267423
764	0.90290550	0.90756300	0.90946790	-2.477349
772	0.03001808	0.02575992	0.02093909	2.824452
815	0.14693120	0.12092930	0.11909650	3.209470

	diff_results\$permutation_p[diff_list_perm]
66	0
129	0
193	0
245	0
432	0
454	0
460	0
601	0
660	0
690	0
764	0
772	0
815	0

```
> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t)
> print(sig_results_boot)
```

	IlmnID	case1	case2	control1	control2	case3
76	cg00065408	0.03952223	0.03967472	0.04799694	0.04929252	0.04064262
110	cg00098239	0.02698720	0.02142180	0.01856646	0.01934917	0.02510008
129	cg00121158	0.03045297	0.02728770	0.03573820	0.03316130	0.02853104
593	cg00568792	0.07330291	0.08046910	0.07976878	0.08124407	0.07145583
887	cg00862290	0.43640520	0.50563130	0.60786800	0.56325950	0.50259740

  

	case4	control3	control4	diff_results\$ordfit_t[diff_list_boot]
76	0.03622954	0.04778213	0.04327211	-3.115264
110	0.02291811	0.01784160	0.02109290	1.976098
129	0.02993313	0.03318921	0.03345607	-2.118418
593	0.07302756	0.08203244	0.08172888	-2.328352
887	0.51368550	0.56646700	0.54552980	-3.935409

  

	diff_results\$bootstrap_p[diff_list_boot]
76	0
110	0
129	0
593	0
887	0