

# Package ‘calm’

October 16, 2020

**Type** Package

**Title** Covariate Assisted Large-scale Multiple testing

**Version** 1.2.0

**Description** Statistical methods for multiple testing with covariate information. Traditional multiple testing methods only consider a list of test statistics, such as p-values. Our methods incorporate the auxiliary information, such as the lengths of gene coding regions or the minor allele frequencies of SNPs, to improve power.

**License** GPL (>=2)

**Encoding** UTF-8

**LazyData** false

**Imports** mgcv, stats, graphics

**Suggests** knitr, rmarkdown

**VignetteBuilder** knitr

**biocViews** Bayesian, DifferentialExpression, GeneExpression, Regression, Microarray, Sequencing, RNASeq, MultipleComparison, Genetics, ImmunoOncology, Metabolomics, Proteomics, Transcriptomics

**RoxygenNote** 6.1.1

**BugReports** <https://github.com/k22liang/calm/issues>

**git\_url** <https://git.bioconductor.org/packages/calm>

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**Description**

Statistical methods for multiple testing with covariate information.

**Details**

Package:	calm
Type:	Package
Version:	0.9.0
Date:	2019-06-22
License:	GPL (>= 2)
LazyLoad:	yes

**Author(s)**

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**References**

Liang, K (2019) *Empirical Bayes analysis of RNA sequencing experiments with auxiliary information*.

**See Also**

[CLfdr](#)

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CLfdr	<i>Conditional local FDR (CLfdr)</i>
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**Description**

CLfdr returns the local false discovery rate (FDR) conditional on auxiliary covariate information

**Usage**

```
CLfdr(x, y, pval = NULL, pi0.method = "RB", bw.init = NULL,  
      bw = NULL, reltol = 1e-04, n.subsample = NULL, check.gam = FALSE,  
      k.gam = NULL, info = TRUE)
```

**Arguments**

<code>x</code>	covariates, could be a vector of length $m$ or a matrix with $m$ rows.
<code>y</code>	a vector of $z$ -values of length $m$ .
<code>pval</code>	a vector of $p$ -values of length $m$ . The $p$ -values are only used to computed the overall true null proportion when <code>pi0.method="RB"</code> .
<code>pi0.method</code>	method to estimate the overall true null proportion ( $\pi_0$ ). "RB" for the right-boundary procedure (Liang and Nettleton, 2012, JRSSB) or "JC" (Jin and Cai, 2007, JASA).
<code>bw.init</code>	initial values for bandwidth, optional. If not specified, normal-reference rule will be used.
<code>bw</code>	bandwidth values.
<code>reltol</code>	relative tolerance in optim function.
<code>n.subsample</code>	size of the subsample when estimating bandwidth.
<code>check.gam</code>	indicator to perform <code>gam.check</code> function on the nonparametric fit.
<code>k.gam</code>	tuning parameter for <code>mgcv::gam</code> .
<code>info</code>	indicator to print out fitting information.

**Details**

In many multiple testing applications, the auxiliary information is widely available and can be useful. Such information can be summary statistics from a similar experiment or disease, the lengths of gene coding regions, and minor allele frequencies of SNPs.

`y` is a vector of  $m$   $z$ -values, one of each hypothesis under test. The  $z$ -values follow  $N(0,1)$  if their corresponding null hypotheses are true. Other types of test statistics, such as  $t$ -statistics and  $p$ -values can be transformed to  $z$ -values. In practice, if the distribution of  $z$ -values is far from  $N(0,1)$ , recentering and rescaling of the  $z$ -values may be necessary.

`x` contains auxiliary covariate information. For a single covariate, `x` should be a vector of length  $m$ . For multiple covariates, `x` should be a matrix with  $m$  rows. The covariates can be either continuous or ordered.

`pi0.method` specifies the method used to estimate the overall true null proportion. If the  $z$ -values are generated from the normal means model, the "JC" method from Jin and Cai (2007) JASA can be a good candidate. Otherwise, the right-boundary procedure ("RB", Liang and Nettleton, 2012, JRSSB) is used.

`bw` are bandwidth values for estimating local alternative density. Suppose there are  $p$  covariates, then `bw` should be a vector of  $p+1$  positive numerical values. By default, these bandwidth values are chosen by cross-validation to minimize a certain error measure. However, finding the optimal bandwidth values by cross-validation can be computationally intensive, especially when  $p$  is not small. If good estimates of bandwidth values are available, for example, from the analysis of a similar dataset, the bandwidth values can be specified explicitly to save time.

`reltol` specifies the relative convergence tolerance when choosing the bandwidth values (`bw`). It will be passed on to `stats::optim()`. For most analyses, the default value of  $1e-4$  provides reasonably good results. A smaller value such as  $1e-5$  or  $1e-6$  could be used for further improvement at the cost of more computation time.

**Value**

<code>fdr</code>	a vector of local FDR estimates. <code>fdr[i]</code> is the posterior probability of the <i>i</i> th null hypothesis is true given all the data. <code>1-fdr[i]</code> is the posterior probability of being a signal (the corresponding null hypothesis is false).
<code>FDR</code>	a vector of FDR values (q-values), which can be used to control FDR at a certain level by thresholding the FDR values.
<code>pi0</code>	a vector of true null probability estimates. This contains the prior probabilities of being null.
<code>bw</code>	a vector of bandwidths for conditional alternative density estimation
<code>fit.gam</code>	an object of <code>mgcv::gam</code>

**Author(s)**

Kun Liang, <kun.liang@uwaterloo.ca>

**References**

Liang (2019), Empirical Bayes analysis of RNA sequencing experiments with auxiliary information, to appear in *Annals of Applied Statistics*

**Examples**

```
data(pso)
ind.nm <- is.na(pso$tval_mic)
x <- pso$len_gene[ind.nm]
# normalize covariate
x <- rank(x)/length(x)
y <- pso$zval[ind.nm]
# assign names to the z-values helps to give names to the output variables
names(y) <- row.names(pso)[ind.nm]

fit.nm <- CLfdr(x=x, y=y)
fit.nm$fdr[1:5]
```

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EstFDR

*FDR estimation*

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**Description**

False discovery rate (FDR) estimation from local FDR

**Usage**

```
EstFDR(fdr)
```

**Arguments**

`fdr` vector of local FDR

**Value**

the estimate of the FDR

**Examples**

```
lfdr <- c(runif(900), rbeta(100, 1, 10))
FDR <- EstFDR(lfdr)
sum(FDR<0.05)
```

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EstNullProp_RB	<i>Right-boundary procedure</i>
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**Description**

True null proportion ( $\pi_0$ ) estimator of Liang and Nettleton (2012), JRSSB

**Usage**

```
EstNullProp_RB(pval, lambda.vec = 0.05 * seq_len(19))
```

**Arguments**

**pval** vector of p-values  
**lambda.vec** vector of lambda candidates (excluding 0 and 1)

**Value**

the estimate of the overall true null proportion

**Examples**

```
pval <- c(runif(900), rbeta(100, 1, 10))
EstNullProp_RB(pval)
```

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pso	<i>Psoriasis RNA-seq dataset</i>
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**Description**

A dataset containing the test statistics to analyze an RNA-seq study of psoriasis.

**Usage**

```
pso
```

**Format**

A dataset with the following vectors:

**zval** 16490 z-values of genes with matching microarray data

**len\_gene** 16490 gene coding region length for zval

**tval\_mic** 16490 matching microarray t-statistics

**Source**

Liang (2019), Empirical Bayes analysis of RNA sequencing experiments with auxiliary information, to appear in *Annals of Applied Statistics*;

**Examples**

```
data(pso)
dim(pso)
# total number of genes without matching microarray data
sum(is.na(pso$tval_mic))
```

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